



Inhibition of vagally mediated gastric acid secretion by activation of central prostanoid EP₃ receptors in urethane-anaesthetized rats

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1 We studied the effects of intracerebroventricular (i.c.v.) administration of prostanoid EP receptor ligands on vagally stimulated gastric acid secretion in rats anaesthetized with urethane.

2 Administration of misoprostol (EP₃/EP₂ receptor agonist) and sulprostone (EP₃/EP₁ receptor agonist) reduced vagally mediated gastric acid secretion in a dose-dependent manner (0.1, 0.3 and 1.0 nmol per animal). Butaprost (EP₂ receptor agonist) (0.3 and 3.0 nmol per animal) was without effect. 17-Phenyl- ω -trilor PGE₂ (EP₁/EP₃ receptor agonist) attenuated vagally mediated gastric acid secretion only at its highest dose (1.0 nmol per animal); this antisecretory effect was not prevented by pretreatment with SC-19220 (selective EP₁ receptor antagonist) (20 nmol per animal, i.c.v.).

3 The potency of these test agents in attenuation of vagally mediated gastric acid secretion was as follows: misoprostol \geq sulprostone $>>$ 17-phenyl- ω -trilor PGE₂ $>>>$ butaprost. These results suggest that activation of central prostanoid EP₃ receptors induces inhibition of vagally mediated gastric acid secretion in rats.

Keywords: Central nervous system; EP₃ receptor; gastric acid secretion

Introduction

Prostaglandins are produced in the central nervous system of mammals (Wolfe, 1982) and are implicated in central regulation of a variety of functions, including body temperature (Cocconi *et al.*, 1988), hypothalamic and pituitary hormone release (Behrman, 1979), some behavioural activities (Johnson *et al.*, 1993) and the control of gastric functions (Saperas *et al.*, 1991; Sautereau *et al.*, 1991). Receptors activated by prostaglandin E₂ (PGE₂) are pharmacologically divided into at least three subtypes (EP₁, EP₂ and EP₃). Many prostanoid ligands with affinity for the EP receptors have been produced, but none shows absolute selectivity for each of the EP receptor subtypes (Coleman *et al.*, 1994). Recently, we reported that i.c.v. administered sulprostone (EP₃/EP₁ receptor agonist) and misoprostol (EP₃/EP₂ receptor agonist) elevated plasma nor-adrenaline levels in urethane-anaesthetized rats, but butaprost (EP₂ receptor agonist) had no effect (Yokotani *et al.*, 1995). From these results, we concluded that central prostanoid EP₃ receptors affect the central sympathetic outflow in rats.

With regard to gastric functions, we previously reported that i.c.v. administered PGE₂ inhibited the vagally stimulated gastric acid secretion and this antisecretory effect was abolished both by bilateral splanchnectomy and by pretreatment with phentolamine (Yokotani *et al.*, 1988). This evidence suggests that activation of central prostanoid EP receptors induces a sympathetic outflow, thereby inhibiting the vagally mediated gastric acid secretion by activation of α -adrenoceptors in the stomach. However, the EP receptor subtypes mediating this effect have not been characterized. In the present study, therefore, we made an attempt to clarify central EP receptor subtypes mediating the inhibition of vagally mediated gastric acid secretion using several kinds of EP receptor ligands in urethane-anaesthetized rats.

Methods

Procedures

Male Wistar rats weighing 350 to 400 g were deprived of food for 16 h but were allowed free access to tap water. Details of

the experimental procedures were as described in our previous papers (Yokotani *et al.*, 1988). Briefly, under urethane anaesthesia (1.2 g kg⁻¹, i.p.), the oesophagus was ligated and the trachea was cannulated through a cervical incision. Bilateral vagus nerves were carefully separated from the carotid arteries and cut at the cervical portion. The peripheral end of the left side vagus nerve was placed on platinum ring electrodes and covered with cotton wool soaked in paraffin oil. The cervical incision was then sutured. The femoral vein was cannulated for infusion of saline (1.5 ml h⁻¹). The abdomen was opened by a midline incision and a round-tip polyethylene cannula (3.5 cm in length and 0.4 cm in diameter) inserted into the stomach via an incision in the duodenum. The cannula was held in place by two ligatures around the duodenum, one at the rostral site and the other at the caudal site of the duodenal incision, and the abdominal incision was sutured. After these procedures, the animal was placed in a stereotaxic apparatus.

Measurement of gastric acid secretion

One hour was allowed to elapse before the start of each experiment, for stabilization of the basal acid secretion. Two ml of gastric solution prewarmed to 38°C was instilled and replaced at intervals of 15 min. The solution consisted of a 1/5 (v/v) mixture of glycine and mannitol adjusted to 300 mOsmol and pH 3.5 by the addition of 0.1 N HCl, according to Blair *et al.* (1975). After two 15 min collections at the basal state, gastric acid secretion was elicited by continuous electrical stimulation of the left vagus nerve using the electronic stimulator (Nihon-Kohden SEN-7103, Tokyo, Japan) and isolator (Nihon-Kohden SS-102J). Stimulation was with square-wave pulses of 0.5 ms duration, at 3 Hz and supramaximal intensity (1 mA). Gastric acid secretion was determined by titration with 0.01 N NaOH to pH 7.0 and expressed as $\mu\text{Eq } 15 \text{ min}^{-1}$.

I.c.v. administration of prostanoid EP receptor agonists and antagonists

For i.c.v. administration of prostanoid EP receptor agonists (sulprostone, misoprostol, butaprost and 17-phenyl- ω -trilor PGE₂) or their vehicles in all experiments, a stainless steel cannula (0.35 mm outer diameter) was inserted into the lateral cerebral ventricle at co-ordinates AP -0.8 mm from the

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bregma, L 1.5 mm from the midline, H 4.0 mm below the surface of the brain according to the rat brain atlas of Paxinos & Watson (1986). Vehicle or prostanoid EP receptor agonist dissolved in saline was slowly injected into the right lateral ventricle in a volume of 10 μ l with a 50 μ l Hamilton syringe 60 min after the start of the vagal stimulation. In some experiments, SC-19220 was administered i.c.v. in a volume of 10 μ l 30 min before the beginning of vagal stimulation.

Compounds

The following drugs were used: 17-phenyl- ω -trinor PGE₂ was purchased from Cayman Chemicals, U.S.A. The following compounds were gifts which we gratefully acknowledge: butaprost from Bayer, U.K.; sulprostone from Schering AG, Germany; misoprostol and SC-19220 (1-acetyl-2-(8-chloro-10, 11-dihydrodibenz (b,f) (1,4) oxazepine-10-carbonyl)-hydrazine) from G.D. Searle, U.S.A. Prostanoid EP receptor agonists were stored at -20°C in 99% ethanol and diluted with saline to 0.5% ethanol concentration whenever they were used (saline containing 0.5% of ethanol was used as vehicle for control experiments). SC-19220 was dissolved in saline containing 5% dimethylsulphoxide.

Treatment of data and statistics

Because the absolute values of acid secretion varied with individual animals, the effects of prostanoid EP receptor agonists on gastric acid secretion were expressed as percentages of the value obtained immediately before i.c.v. administration of these ligands. Results were expressed as the mean \pm s.e.mean. Statistical analysis was carried out using the Bonferroni method for comparing a control to all other means after one-way analysis of variance (ANOVA) in Figures 1 and 2 and Student's unpaired *t* test in Figure 3. *P* values of less than 0.05 were taken to indicate significant differences.

Results

Effects of misoprostol (EP₃/EP₂ receptor agonist) and sulprostone (EP₃/EP₁ receptor agonist) on the gastric acid secretion induced by vagus nerve stimulation

Continuous electrical stimulation of the vagus nerve at 3 Hz evoked an increase in gastric acid secretion which lasted more than 60 min (Figure 1a). Acid secretion at 60 min after the intracerebroventricular (i.c.v.) administration of vehicle was $102.4 \pm 6.5\%$ of the preadministered values at 0 min ($n=4$). The actual values of the evoked gastric acid secretion at 0 min were $92.6 \pm 4.9 \mu\text{Eq } 15 \text{ min}^{-1}$ ($n=44$).

Misoprostol (0.1, 0.3 and 1.0 nmol per animal, i.c.v.) dose-dependently reduced the vagally stimulated gastric acid secretion and these inhibitory effects lasted for more than 60 min after its administration (Figure 1a). Acid secretion at 45 min after administration of misoprostol (0.1, 0.3 and 1.0 nmol per animal, i.c.v.) was 57.8 ± 5.7 ($n=4$), $43.7 \pm 5.4\%$ ($n=4$), and $37.3 \pm 3.9\%$ ($n=4$) of the preadministered values, respectively. On the other hand, intravenous administration of misoprostol (1.0 nmol per animal) was without effect on the vagally stimulated gastric acid output, indicating that the sites of action of the i.c.v. administered misoprostol are probably within the brain but not in peripheral organs after its leakage into systemic circulation (data not shown).

Sulprostone (0.1, 0.3 and 1.0 nmol per animal, i.c.v.) also dose-dependently reduced the vagally stimulated gastric acid secretion and these inhibitory effects lasted over 60 min after its administration (Figure 1b). Acid secretion at 45 min after administration of sulprostone (0.1, 0.3 and 1.0 nmol per animal, i.c.v.) was $61.7 \pm 6.3\%$ ($n=4$), $41.1 \pm 9.2\%$ ($n=4$), and $41.8 \pm 5.1\%$ ($n=4$) of the preadministered values, respectively.

Effects of butaprost (EP₂ receptor agonist) and 17-phenyl- ω -trinor-PGE₂ (EP₁/EP₃ receptor agonist) on gastric acid secretion induced by vagus nerve stimulation

Butaprost (0.3 and 3.0 nmol per animal, i.c.v.) did not significantly affect the vagally stimulated gastric acid secretion (Figure 2a).

17-Phenyl- ω -trinor-PGE₂ (1.0 nmol per animal, i.c.v.) significantly ($P<0.05$) reduced the vagally stimulated gastric acid secretion and this inhibitory effect lasted for 45 min after its administration (Figure 2b). A lower dose of this agent (0.3 nmol per animal, i.c.v.) was without effect.

Effects of SC-19220 (selective EP₁ receptor antagonist) on 17-phenyl- ω -trinor-PGE₂-induced inhibition of gastric acid secretion

After animals had been treated with SC-19220 (20 nmol per animal, i.c.v.) at -90 min, vagal stimulation was initiated at -60 min. I.c.v. administration of SC-19220 had no effect on either the basal or vagally-induced gastric acid secretion (Figure 3). The actual values of the evoked gastric acid secretion at 0 min in SC-19220-treated animals were $84.1 \pm 6.2 \mu\text{Eq } 15 \text{ min}^{-1}$ ($n=8$). 17-Phenyl- ω -trinor-PGE₂ (1.0 nmol per animal) or vehicle were subsequently adminis-

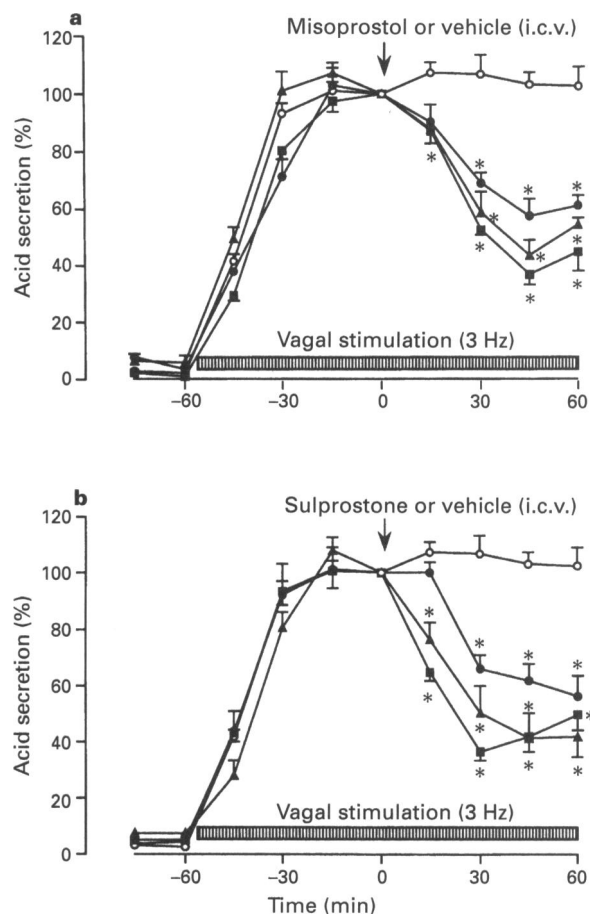


Figure 1 Effects of intracerebroventricularly (i.c.v.) administered misoprostol and sulprostone on vagally stimulated gastric acid secretion. Arrow indicates i.c.v. administration of test substances. In (a), vehicle (\circ), $n=4$; misoprostol 0.1 nmol per animal (\bullet), $n=4$; misoprostol 0.3 nmol per animal (\blacktriangle), $n=4$; misoprostol 1.0 nmol per animal (\blacksquare), $n=4$. In (b), vehicle (\circ) (as in a), $n=4$; sulprostone 0.1 nmol per animal (\bullet), $n=4$; sulprostone 0.3 nmol per animal (\blacktriangle), $n=4$; sulprostone 1.0 nmol per animal (\blacksquare), $n=4$. Each point represents the mean \pm s.e.mean. *Significantly different ($P<0.05$) from vehicle-treated control.

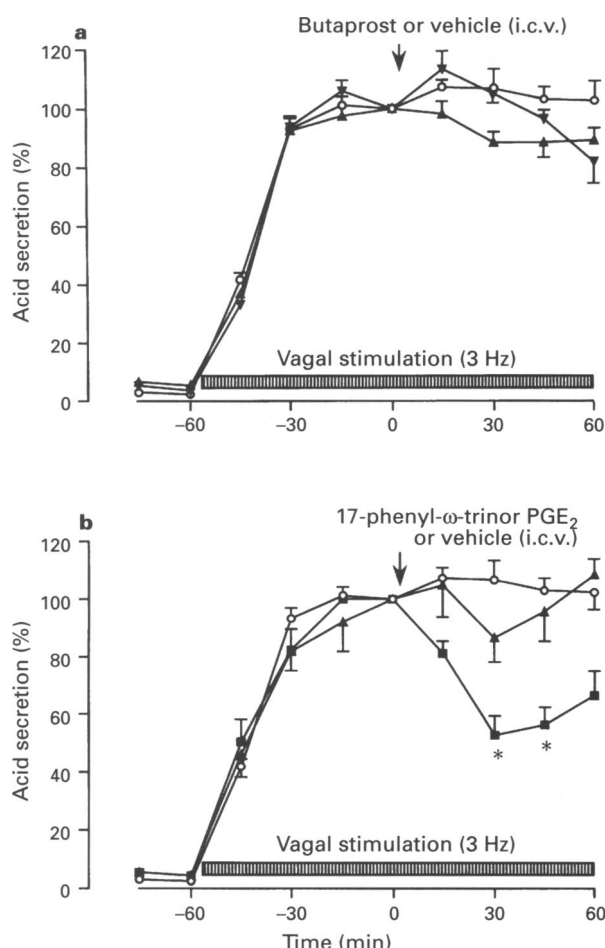


Figure 2 Effects of i.c.v. administered butaprost and 17-phenyl- ω -trinor-PGE₂ on vagally stimulated gastric acid secretion. Arrow indicates i.c.v. administration of test substances. In (a), vehicle (○), $n=4$ (as in Figure 1); butaprost 0.3 nmol per animal (▲), $n=4$; butaprost 3.0 nmol per animal (▼), $n=4$. In (b), vehicle (○) (as in Figure 1); 17-phenyl- ω -trinor-PGE₂ 0.3 nmol per animal (▲), $n=4$; 17-phenyl- ω -trinor-PGE₂ 1.0 nmol per animal (■), $n=4$. *Significantly different ($P<0.05$) from vehicle-treated control. Other conditions were the same as those in Figure 1.

tered i.c.v. (at 0 min); 17-phenyl- ω -trinor-PGE₂ reduced the vagally stimulated gastric acid secretion. Acid secretion at 45 min after i.c.v. administration of 17-phenyl- ω -trinor-PGE₂ (1.0 nmol per animal) was $39.0 \pm 5.6\%$ of the preadministered values.

Discussion

Puurunen (1983; 1984) demonstrated that i.c.v. administered PGE₂ (3–10 μ g per animal) decreased gastric acid secretion in conscious, pylorus-ligated rats. Since this inhibition was prevented by hypophysectomy or intravenous administration of a vasopressin antagonist, he concluded that the central antisecretory action of PGE₂ was due to the release of vasopressin from the pituitary gland. In our previous report, however, i.c.v. but not i.v. administered small doses of PGE₂ (0.05–0.5 μ g per animal) also effectively inhibited the vagally stimulated gastric acid secretion in urethane-anaesthetized rats (Yokotani *et al.*, 1988). This PGE₂-induced antisecretory effect was abolished both by bilateral splanchnectomy and by pre-treatment with phentolamine, an α -adrenoceptor blocking agent. Activation of α -adrenoceptors in the stomach inhibits the vagally stimulated gastric acid secretion by reducing acetylcholine release from parasympathetic nerve terminals in the

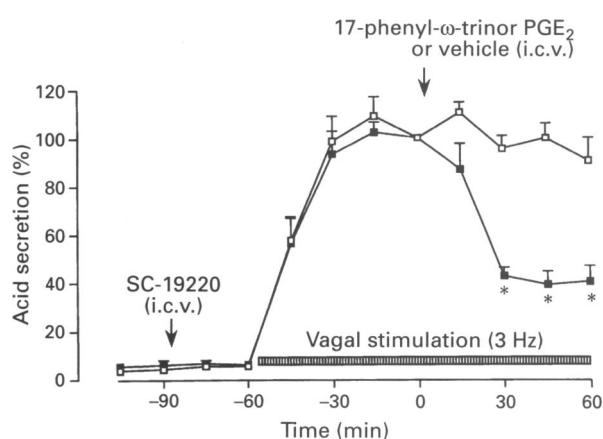


Figure 3 Effects of SC-19220 on the 17-phenyl- ω -trinor-PGE₂-induced inhibition of gastric acid secretion. The first arrow (at -90 min) indicates the administration of SC-19220 (20 nmol per animal, i.c.v.). The second arrow (at 0 min) indicates the administration of 17-phenyl- ω -trinor-PGE₂ (1.0 nmol per animal, i.c.v.) or vehicle alone (i.c.v.). Vehicle (□), $n=4$; 17-phenyl- ω -trinor-PGE₂ 1.0 nmol per animal (■), $n=4$. *Significantly different ($P<0.05$) from SC-19220 plus vehicle-treated control. Other conditions were the same as in Figures 1 and 2.

stomach (Yokotani *et al.*, 1984; 1993). This evidence suggests that centrally administered PGE₂ inhibits gastric acid secretion by activation of central sympathetic outflow. Furthermore, vasopressin released from the pituitary gland with a relatively large dose of PGE₂ also inhibits gastric acid secretion as suggested by Puurunen (1983, 1984).

In the present experiments, we used various agonists selective for EP receptor subtypes to characterize the central EP receptor which mediates inhibition of gastric acid secretion. Misoprostol is a potent EP₃ receptor agonist and also has moderate EP₂ and weak EP₁ receptor agonist activities (agonist potencies: EP₃>EP₂>>EP₁) (Lawrence *et al.*, 1992; Mantelli *et al.*, 1991). Sulprostone was first identified as a potent EP₁ receptor agonist, but it is more potent at EP₃ receptors (agonist potencies: EP₃>EP₁>>EP₂) (Coleman *et al.*, 1987a,b; Bunce *et al.*, 1990). Butaprost behaves as a selective EP₂ receptor agonist with relatively low potency (agonist potencies: EP₂>>>EP₁) (Gardiner, 1986). In the present experiments, misoprostol and sulprostone (0.1, 0.3 and 1.0 nmol per animal, i.c.v.) both effectively reduced vagally mediated gastric acid secretion in a dose-dependent manner. However, butaprost was ineffective on the vagally mediated gastric acid secretion. These results suggest that EP₃ receptors in the brain are probably involved in the PGE₂-induced antisecretory effects. However, there was a possibility that central EP₁ receptors are also involved in these antisecretory effects.

We finally examined the role of EP₁ receptors in the PGE₂-induced antisecretory effect. 17-Phenyl- ω -trinor PGE₂ is an EP₁ receptor agonist, with some weaker EP₃ agonist activity (agonist potencies: EP₁>EP₃>>EP₂) (Dong *et al.*, 1986; Lawrence *et al.*, 1989; 1992). In the present study, the dose of 17-phenyl- ω -trinor PGE₂ (1.0 nmol per animal, i.c.v.), sufficient to reduce the vagally mediated gastric acid secretion, was relatively large. Furthermore, SC-19220, a highly selective and competitive albeit rather weak EP₁ receptor antagonist (Santer, 1969; Kennedy *et al.*, 1982; Coleman *et al.*, 1985), failed to attenuate 17-phenyl- ω -trinor PGE₂-induced inhibition of gastric acid secretion at a dose of 20 nmol per animal (i.c.v.). This dose of SC-19220 is probably sufficient to block central EP₁ receptors (Ferreira *et al.*, 1978; Kandasamy & Williams, 1982). The present results therefore suggest that an antisecretory effect of a large dose of 17-phenyl- ω -trinor PGE₂ (1.0 nmol per animal, i.c.v.) is produced by activation of central EP₃-receptors.

In the central nervous system, EP₃ receptor mRNA is highly expressed in brain regions such as the preoptic area, hypothalamus, locus coeruleus and raphe nuclei (Sugimoto *et al.*, 1994). Administration of PGE₂ into specific brain areas of the hypothalamus such as ventromedial hypothalamus and paraventricular nucleus of hypothalamus has been shown to inhibit stimulated gastric acid secretion (Barocelli *et al.*, 1991). The paraventricular nucleus of the hypothalamus is a major site sending signals to the spinal sympathetic preganglionic neurones (Swanson & Sawchenko, 1983). Recently we reported that activation of central EP₃ receptors elevates plasma noradrenaline levels (Yokotani *et al.*, 1995). It is interesting to note that the potencies of misoprostol, sulprostone and 17-phenyl- ω -trilor PGE₂ in elevating plasma noradrenaline levels were almost the same as those for inhibition of

gastric acid secretion. Furthermore, misoprostol- and sulprostone-induced antisecretory effects were abolished by pre-treatment with phentolamine (5 mg kg⁻¹, i.m.) (data not shown). It is therefore likely that the PGE₂-induced antisecretory effect is mediated by central prostanoid EP₃ receptors through activation of central sympathetic outflow in urethane-anaesthetized rats.

In summary, we demonstrate here that EP₃-receptors in the rat brain are involved in the PGE₂-induced antisecretory effects on vagally mediated gastric acid secretion.

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References

- BLAIR, E.L., GRUND, E.R., REED, J.D., SANDERS, D.J., SANGER, G. & SHAW, B. (1975). The effect of sympathetic nerve stimulation on serum gastrin, gastric acid secretion and mucosal blood flow responses to meat extract stimulation in anaesthetized cats. *J. Physiol.*, **253**, 493–504.
- BAROCELLI, E., IMPICCIATORE, M., SEATON, J., CONTER, R. & KAUFFMAN, G. (1991). Localization of central prostaglandin E₂ antisecretory effects. *Gastroenterology*, **100**, 320–327.
- BEHRMAN, H.R. (1979). Prostaglandins in hypothalamo-pituitary and ovarian function. *Annu. Rev. Physiol.*, **41**, 685–700.
- BUNCE, K.T., CLAYTON, N.M., COLEMAN, R.A., COLLINGTON, E.W., FINCH, H., HUMPHRAY, J.M., HUMPHREY, P.P.A., REEVES, J.J., SHELDRICK, R.L.G. & STABLES, R. (1990). GR63799X - a novel prostanoid with selectivity for EP₃-receptors. *Adv. Prostaglandin Thromboxane Leukot. Res.*, **21**, 379–382.
- COCEANI, F., LEES, J. & BISHAI, I. (1988). Further evidence implicating prostaglandin E₂ in the genesis of pyrogen fever. *Am. J. Physiol.*, **254**, R463–R469.
- COLEMAN, R.A., KENNEDY, I. & SHELDRICK, R.L.G. (1985). AH6809, a prostanoid EP₁ receptor blocking drug. *Br. J. Pharmacol.*, **85**, 273P.
- COLEMAN, R.A., KENNEDY, I. & SHELDRICK, R.L.G. (1987a). Evidence for the existence of three subtypes of PGE₂ sensitive (EP) receptors in smooth muscle. *Br. J. Pharmacol.*, **91**, 323P.
- COLEMAN, R.A., KENNEDY, I., SHELDRICK, R.L.G. & TOLOWINSKA, I.Y. (1987b). Further evidence for the existence of three subtypes of PGE₂-sensitive (EP-) receptors. *Br. J. Pharmacol.*, **91**, 407P.
- COLEMAN, R.A., SMITH, W.L. & NARUMIYA, S. (1994). Classification of prostanoid receptors: properties, distribution, and structure of the receptors and their subtypes. *Pharmacol. Rev.*, **46**, 205–229.
- DONG, Y.J., JONES, R.L. & WILSON, N.H. (1986). Prostaglandin E receptor subtypes in smooth muscle: agonist activities of stable prostacyclin analogues. *Br. J. Pharmacol.*, **87**, 97–107.
- FERREIRA, S.H., LORENZETTI, B.B. & CORRÊA, F.M.A. (1978). Central and peripheral antialgesic action of aspirin-like drugs. *Eur. J. Pharmacol.*, **53**, 39–48.
- GARDINER, P.J. (1986). Characterization of prostanoid relaxant/inhibitory receptors (Ψ) using a highly selective agonist, TR 4979. *Br. J. Pharmacol.*, **87**, 45–56.
- JOHNSON, R.W., CURTIS, S.E., DANTZER, R. & KELLEY, K.W. (1993). Central and peripheral prostaglandins are involved in sickness behaviour in birds. *Physiol. Behav.*, **53**, 127–131.
- KANDASAMY, S.B. & WILLIAMS, B.A. (1982). Prostacyclin-induced hyperthermia: Implication of a protein mediator. *Neuropharmacology*, **21**, 1065–1072.
- KENNEDY, I., COLEMAN, R.A., HUMPHREY, P.P.A., LEVY, G.P. & LUMLEY, P. (1982). Studies on the characterisation of prostanoid receptors: a proposed classification. *Prostaglandins*, **24**, 667–689.
- LAWRENCE, R.A., JONES, R.L. & WILSON, N.H. (1989). Relaxant properties of prostaglandin E analogues on rabbit jugular vein. *Br. J. Pharmacol.*, **98**, 796P.
- LAWRENCE, R.A., JONES, R.L. & WILSON, N.H. (1992). Characterization of receptors involved in the direct and indirect actions of prostaglandins E and I on the guinea-pig ileum. *Br. J. Pharmacol.*, **105**, 271–278.
- MANTELLI, L., AMRINI, S., RUBINO, A. & LEDDA, F. (1991). Prejunctional prostanoid receptors on cardiac adrenergic terminals belong to the EP₃ subtype. *Br. J. Pharmacol.*, **102**, 573–576.
- PAXINOS, G. & WATSON, C. (ed.) (1986). *The Rat Brain in Stereotaxic Coordinates*. Boston: Academic Press, Inc.
- PUURUNEN, J. (1983). Central nervous system effects of arachidonic acid, PGE₂, PGF_{2 α} , PGD₂ and PGI₂ on gastric secretion in the rat. *Br. J. Pharmacol.*, **80**, 255–262.
- PUURUNEN, J. (1984). Central gastric antisecretory action of prostaglandin E₂: The involvement of the pituitary gland. *J. Pharmacol. Exp. Ther.*, **231**, 713–716.
- SANNER, J.H. (1969). Antagonism of prostaglandin E₂ by 1-acetyl-2-(8-chloro-10, 11-dihydrobenz (b,f) (1,4) oxazepine-10-carbonyl) hydrazine (SC-19220). *Arch. Int. Pharmacodyn. Ther.*, **180**, 46–56.
- SAPERAS, E., KAUFFMAN, G. & TACHÉ, Y. (1991). Role of central prostaglandin E₂ in the regulation of gastric acid secretion in the rat. *Eur. J. Pharmacol.*, **209**, 1–7.
- SAUTEREAU, D., CHICAU, C.M., TSOCAS, A. & ROZE, C. (1991). Central and peripheral effects of prostaglandin E₂ and enprostil on gastric acid secretion in the rats. *Eur. J. Pharmacol.*, **195**, 217–224.
- SUGIMOTO, Y., SHIGEMOTO, R., NAMBA, T., NEGISHI, M., MISUNO, N., NARUMIYA, S. & ICHIKAWA, A. (1994). Distribution of the messenger RNA for the prostaglandin E receptor subtype EP₃ in the mouse nervous system. *Neuroscience*, **62**, 919–928.
- SWANSON, L.W. & SAWCHENKO, P.E. (1983). Hypothalamic integration: Organization of the paraventricular and supraoptic nuclei. *Annu. Rev. Neurosci.*, **6**, 269–324.
- WOLFE, L.S. (1982). Eicosanoids: Prostaglandins, thromboxanes, leukotrienes, and other derivatives of carbon-20 unsaturated fatty acids. *J. Neurochem.*, **38**, 1–14.
- YOKOTANI, K., MURAMATSU, I. & FUJIWARA, M. (1984). Alpha-1 and alpha-2 type adrenoceptors involved in the inhibitory effect of splanchnic nerves on parasympathetically stimulated gastric acid secretion in rats. *J. Pharmacol. Exp. Ther.*, **229**, 305–310.
- YOKOTANI, K., NISHIHARA, M., MURAKAMI, Y., HASEGAWA, T., OKUMA, Y. & OSUMI, Y. (1995). Elevation of plasma noradrenaline levels in urethane-anaesthetized rats by activation of central prostanoid EP₃ receptors. *Br. J. Pharmacol.*, **115**, 672–676.
- YOKOTANI, K., OKUMA, Y., NAKAMURA, K. & OSUMI, Y. (1993). Release of endogenous acetylcholine from a vascularly perfused rat stomach in vitro; Inhibition by M₃ muscarinic autoreceptors and alpha-2 adrenoceptors. *J. Pharmacol. Exp. Ther.*, **266**, 1190–1195.
- YOKOTANI, K., YOKOTANI, K., OKUMA, Y. & OSUMI, Y. (1988). Sympathoadrenomedullary system mediation of the prostaglandin E₂-induced central inhibition of gastric acid output in rats. *J. Pharmacol. Exp. Ther.*, **244**, 335–340.

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